

BIRCH, STEWART, KOLASCH & BIRCH, LLP

INTELLECTUAL PROPERTY LAW

8110 GATEHOUSE ROAD

SUITE 500 EAST

FALLS CHURCH, VA 22042-1210

USA

(703) 205-8000

FAX: (703) 205-8050

(703) 698-8590 (G IV)

e-mail: mailroom@bskb.com

web: http://www.bskb.com

CALIFORNIA OFFICE

650 TOWN CENTER DRIVE

SUITE 1120

COSTA MESA, CA 92626-7125

(714) 708-8555

FAX: (714) 708-8565

GARY D. YACURA
THOMAS S. AUCHTERLOE
MICHAEL R. CAMMARATA
JAMES T. ELLER, JR.
SCOTT L. LOWE
MARY ANN CAPRIA
MARK J. NUEL, PH.D.
ROBERT V. RACUNAS
DARIN E. BARTHOLOMEW*
D. RICHARD ANDERSON
PAUL C. LEWIS
W. KARL RENNER

REG. PATENT AGENTS
FREDERICK R. HANDREN
ANDREW J. TELESZ, JR.
MARYANNE LIOTTA, PH.D.
MAKI HATSUMI
MIKE S. RYU
CRAIG A. McROBBIE
GARTH M. DAHLEN, PH.D.
LAURA C. LUTZ
ROBERT E. GOOZNER, PH.D.
HYUNG N. SOHN
MATTHEW J. LATTIG
ALAN PEDERSEN-GILES

HERRELL C. BIRCH
RAYMOND C. STEWART
JOSEPH A. KOLASCH
JAMES M. SLATTERY
BERNARD L. SWEENEY*
MICHAEL K. MUTTER
CHARLES GORENSTEIN
RALPH M. MURPHY, JR.
RONALD R. SVENSSON
MARRY L. CLARK
ANDREW D. MEIKLE
MARC S. WEINER
JOE MCKINNEY MUNCY
ROBERT J. KENNEY
C. JOSEPH FARACI
DONALD J. DALEY
JOHN W. BAILEY
JOHN A. CASTELLANO, III
OF COUNSEL
HERBERT M. BIRCH (1905-1996)
ELLIOT A. GOLDBERG*
WILLIAM L. GATES*
EDWARD H. VALANCE
RUPERT J. BRADY (RET.)*
*ADMITTED TO A BAR OTHER THAN VA

Date: August 27, 1999

Docket No.: 2167-0110P

Assistant Commissioner for Patents
Box PATENT APPLICATION
Washington, D.C. 20231

Sir:

Transmitted herewith for filing is the patent application of

Inventor(s): AOKI, Yosuke
SUZUKI, Hideaki; TAKAHASHI, Kiyoshi
KATSURAGI, Hisashi

For: ANTI-HUMAN MEDULLASIN MONOCLONAL ANTIBODY, PROCESS FOR
PRODUCING THE SAME AND IMMUNOASSAY USING THE SAME

Enclosed are:

- ☒ A specification consisting of 18 pages
- ☒ 1 sheet(s) of Formal drawings
- ☒ An assignment of the invention
- ☐ Certified copy of Priority Document(s)
- ☒ Executed Declaration ☒ Original ☐ Photocopy
- ☐ A verified statement to establish small entity status under 37
CFR 1.9 and 37 CFR 1.27
- ☐ Preliminary Amendment
- ☒ Information Disclosure Statement, PTO-1449 and reference(s)

Other _____

The filing fee has been calculated as shown below:

LARGE ENTITY				SMALL ENTITY			
FOR	NO. FILED	NO. EXTRA	RATE	FEE		RATE	FEE
BASIC FEE	*****	*****	*****		or	****	
	*****	*****	*****	\$760.00		****	\$380.00
	*****	*****	*****			****	
TOTAL CLAIMS	5 - 20 =	0	x18 = \$	0.00	or	x 9 = \$	0.00
INDEPENDENT	2 - 3 =	0	x78 = \$	0.00	or	x 39 = \$	0.00
MULTIPLE DEPENDENT CLAIM PRESENTED <u>no</u>				+260 = \$	or	+130 = \$	0.00
				TOTAL \$		TOTAL \$	0.00

X A check in the amount of \$ 800.00 to cover the filing fee and recording fee (if applicable) is enclosed.

_____ Please charge Deposit Account No. 02-2448 in the amount of \$ _____. A triplicate copy of this transmittal form is enclosed.

_____ No fee is enclosed.

If necessary, the Commissioner is hereby authorized in this, concurrent, and future replies, to charge payment or credit any overpayment to Deposit Account No. 02-2448 for any additional fees required under 37 C.F.R. 1.16 or under 37 C.F.R. 1.17; particularly, extension of time fees.

Respectfully submitted,

BIRCH, STEWART, KOLASCH & BIRCH, LLP

By

[Signature] #35,094
GERALD M. MURPHY, JR.

Reg. No. 28,977

P. O. Box 747

Falls Church, Virginia 22040-0747

(703) 205-8000

GMM/sas

SPECIFICATION

ANTI-HUMAN MEDULLASIN MONOCLONAL ANTIBODY,
PROCESS FOR PRODUCING THE SAME AND IMMUNOASSAY
USING THE SAME

BACKGROUND OF THE INVENTION

I. Field of the Invention

The present invention relates to an anti-human medullasin monoclonal antibody, process for producing the same and immunoassay using the same.

II. Description of the Related Art

Medullasin which is a kind of serine proteases occurs in granulocytes and the like, and is thought to widely play important roles in defense mechanism, including expression of inflammation, especially chronic inflammation. The amount of medullasin in granulocytes is increased in advanced stage of a number of chronic inflammatory diseases, and is normalized in remission stage. It is observed in patients suffering from multiple sclerosis that the amount of medullasin is prominently increased several days before the advanced stage and is normalized before remission. Multiple sclerosis is an intractable chronic inflammatory disease which mostly results in death in 10 to 15 years, which is characterized by localized demyelinated lesion in white matter of central nerve system and gliosis, and progresses repeating remission and aggravation. Although the cause of this disease has not been clarified, it is thought that this disease is a kind of autoimmune diseases in which autoantibodies attack the nerve tissue upon stimulation of the immune

system by a virus or a bacterium. Its diagnosis is quite difficult and is now carried out by nuclear magnetic resonance imaging (MRI) or the like. However, MRI or the like requires a very large-scale equipment and high skill in the measuring operation, and is costly. Thus, a simple diagnosis method by which diagnosis of the disease, understanding of the state of the disease and assumption of the consequence can be accomplished is now being developed. For this, methods for measuring the activity of medullasin in granulocytes in the blood are now studied, and a method based on the ability of medullasin to inactivate apo-ornithine transaminase has been developed. However, since measurement of the activity of medullasin in granulocytes by an enzymatic chemical method is very complicated, simple immunoassay was tried and has been developed, which utilizes a polyclonal antibody.

However, the immunoassay for measuring human medullasin using a polyclonal antibody has drawbacks in that the reactivity of the antibody is not necessarily sufficient, the measuring time is long and the measuring operation is somewhat complicated. Thus, an immunoassay utilizing a monoclonal antibody, by which human medullasin can be measured quickly and simply, is demanded.

SUMMARY OF THE INVENTION

The present invention was accomplished under these circumstances and an object of the present invention is to provide a monoclonal antibody which specifically recognizes human medullasin and a method for specifically measuring human medullasin quickly and simply.

The present inventors intensively studied to attain the above-mentioned object and succeeded in obtaining an anti-human medullasin monoclonal antibody by culturing hybridomas prepared by cell fusion

between antibody-producing cells recovered from an animal immunized with human medullasin separated and purified from human granulocytes and myeloma cells according to the method by Köhler and Milstein, and discovered that human medullasin can be measured specifically, quickly and simply by using this antibody, thereby reaching the present invention.

According to the first aspect, the present invention provides an anti-human medullasin monoclonal antibody which specifically recognizes human medullasin.

According to the second aspect, the present invention provides a process for producing an anti-human medullasin monoclonal antibody comprising culturing hybridomas prepared by cell fusion between antibody-producing cells recovered from an animal immunized with human medullasin and myeloma cells, and recovering anti-human medullasin monoclonal antibody which specifically recognizes human medullasin from the culture in which the hybridomas are cultured.

According to the third aspect, the present invention provides an immunoassay comprising immobilizing human medullasin in a test sample by sandwiching the human medullasin between an anti-human medullasin monoclonal antibody immobilized on an insoluble carrier and a labelled anti-human medullasin monoclonal antibody by antigen-antibody reactions to form a complex, and quantifying the label in the complex.

By the present invention, an anti-human medullasin monoclonal antibody which specifically recognizes medullasin that is a kind of serine proteases existing in granulocytes was provided. By utilizing this monoclonal antibody, quick and simple immunoassay for measuring human medullasin was provided, which may be used for hemodiagnosis or the like for chronic inflammatory diseases, especially multiple sclerosis.

BRIEF DESCRIPTION OF THE DRAWINGS

Fig. 1 is a calibration curve for measuring human medullasin, prepared by plotting the absorbance measured by the enzyme immunoassay (Measuring Method I) described in Example 2 as a function of the concentration of the antigen; and

Fig. 2 is a calibration curve for measuring human medullasin, prepared by plotting the absorbance measured by the enzyme immunoassay (Measuring Method II) described in Example 3 as a function of the concentration of the antigen.

DETAILED DESCRIPTION OF THE PREFERRED EMBODIMENTS

The anti-human medullasin monoclonal antibody according to the present invention may be produced by culturing hybridomas in a culture medium, which hybridomas were prepared by cell fusion between antibody-producing cells recovered from an animal immunized with human medullasin extracted from granulocytes separated from the blood of a normal individual and myeloma cells, and recovering the monoclonal antibody from the culture, or by intraperitoneally administering the hybridomas to an animal, proliferating the hybridomas in ascites, and recovering the monoclonal antibody from the ascites.

The hybridomas producing the anti-human medullasin monoclonal antibody according to the present invention may be produced by the so called cell fusion method. That is, the desired monoclonal antibody-producing hybridoma may be obtained by recovering antibody-producing cells from an animal immunized with human medullasin, fusing the antibody-producing cells with myeloma cells, selectively proliferating the obtained hybridomas, screening the antibody-producing hybridomas from the obtained hybridomas, and cloning the selected hybridomas.

Examples of the antibody-producing cells include spleen cells, lymph node cells, B lymphocytes and the like, which are obtained from an animal immunized with human medullasin or a composition or cells containing human medullasin. Examples of the animal to be immunized include mouse, rat, rabbit, goat, sheep, horse and the like. Immunization may be carried out by, for example, subcutaneously, intramuscularly or intraperitoneally administering human medullasin to an animal with a dose of about 1 μ g to 1 mg/time, once or twice per month for 1 to 6 months. Collection of the antibody-producing cells may be carried out 2 to 4 days after the final immunization.

The myeloma cells may be originated from mouse, rat or the like. The antibody-producing cells and the myeloma cells may preferably be originated from the same animal species.

The method of cell fusion may be arbitrarily selected. For example, cell fusion may be carried out by mixing the antibody-producing cells and the myeloma cells in a medium such as Dulbecco's Modified Eagle Medium (DMEM) in the presence of a fusion accelerator such as polyethylene glycol.

After the cell fusion operation, the hybridomas may be selected by appropriately diluting the cells after cell fusion operation with DMEM or the like, centrifuging the resultant, suspending the precipitate in a selection medium such as HAT medium, and culturing the cells therein. Antibody-producing hybridomas are then screened by enzyme immunoassay or the like using the culture supernatant, and the selected hybridoma is cloned by limiting dilution method to obtain the hybridoma producing the anti-human medullasin monoclonal antibody according to the present invention.

The monoclonal antibody according to the present invention may be produced by culturing the thus obtained antibody-producing hybridoma in an appropriate culture medium or in an animal, and recovering the monoclonal antibody from the culture. To produce a large amount of monoclonal antibody, the method in which the hybridomas are intraperitoneally administered to an animal of the same species as the donor of the myeloma cells, the monoclonal antibody according to the present invention is accumulated in the ascites, and the monoclonal antibody is recovered from the ascites, is preferred. Before the intraperitoneal administration of the antibody-producing hybridomas, it is preferred to administer a mineral oil such as pristane.

The monoclonal antibody according to the present invention may be separated from the culture or ascites by fractionation with ammonium sulfate, anion-exchange column chromatography or column chromatography using protein A, G or the like, which is usually employed for purification of IgG.

Four kinds of anti-human medullasin monoclonal antibodies according to the present invention, designated 3F03, 3G03, 2E04 and 1G12, have been obtained. All of these monoclonal antibodies belong to immunoglobulin class IgG, subclass IgG₁, and specifically react with human medullasin which is the corresponding antigen.

Therefore, by using the anti-human medullasin monoclonal antibody according to the present invention, human medullasin may be immunologically measured. The immunoassay for measuring human medullasin using the monoclonal antibody according to the present invention may be arbitrarily selected. A competition method in which the antigen in a sample is reacted with the monoclonal antibody

immobilized on an insoluble carrier in the presence of a labeled antigen; and a sandwich method in which the antigen in a sample is reacted with the monoclonal antibody immobilized on an insoluble carrier, and then the monoclonal antibody which is labeled is reacted with the resultant to form an antibody-antigen-antibody sandwich complex, are preferred. In the enzyme immunoassay according to the sandwich method, it is required that the corresponding epitopes of the immobilized antibody and of the labeled antibody be different, so that monoclonal antibodies of which corresponding epitopes are different are used in combination for constituting the immunoassay system. However, immunoassay system was successfully prepared by using the monoclonal antibody 2E04 according to the present invention as both the immobilized antibody and as the labeled antibody. This is because human medullasin has epitopes in a single molecule, whose amino acid sequences are identical. Examples of the insoluble carriers used for these measuring methods include beads made of a plastic such as polystyrene, and microplate. Preferred examples of the label used for labeling the antibody include enzymes, fluorescent substances, luminescent substances and radioactive substances. Examples of the enzymes include peroxidase, alkaline phosphatase, β -D-galactosidase and the like. Examples of the fluorescent substances include fluorescein isocyanate, phycobiliprotein and the like. Examples of the luminescent substances include luminols, dioxetanes, acridinium salts and the like. Examples of the radioactive substances include ^{125}I , ^{131}I , ^{111}In , $^{99\text{m}}\text{Tc}$ and the like. In cases where the labeling substance is an enzyme, a substrate, and if necessary, a coloring agent, scintillator, luminescent agent or the like are used. In cases where peroxidase is used as the enzyme, hydrogen peroxide or the like may be

used as the substrate, 2,2'-azinodi[3-ethylbenzthiazoline sulfonic acid] ammonium salt (ABTS), 5-aminosalicylic acid, o-phenylenediamine, 4-aminoantipyrine, 3,3',5,5'-tetramethylbenzidine or the like may be used as the coloring agent, 4-hydroxyphenylacetic acid, 3-(4-hydroxyphenyl)propionic acid or the like may be used as the scintillator, and luminols or the like may be used as the luminescent agent. In cases where alkaline phosphatase is used as the enzyme, 4-nitrophenylphosphate, 4-methylumbelliferyl phosphate, cortisol-21-phosphate or the like may be used as the substrate. In cases where β -D-galactosidase is used as the enzyme, 2-nitrophenyl- β -D-galactoside, 3-(2'-spiroadamantane)-4-methoxy-4(3"- β -D-galactosyloxyphenyl)-1,2-dioxetane (AMPGD) or the like may be used as the substrate.

Examples

The present invention will now be described more concretely by way of reference examples and examples. However, the present invention is not restricted to the examples.

In all examples, all % are by weight.

Reference Example 1

Four hundred milliliters of the blood from a normal individual and 6% dextran (molecular weight: 200,000-300,000) solution in physiological saline were mixed at a ratio of blood:aqueous dextran solution = 2:1, and the resulting mixture was lightly stirred with a glass rod, followed by leaving to stand the resultant at 4-8°C for about 1 hour. The precipitated red blood cells were removed and the obtained supernatant was centrifuged at 15,000 rpm, followed by recovering the precipitate to obtain leukocytes. To the obtained leukocytes, extraction buffer containing 1 mM ethylenediamine tetraacetic acid disodium salt

(EDTA) and 1 mM p-chloromercury benzoic acid (PCMB) in 1 M potassium phosphate buffer (PKB) (pH 7.0) was added and the resulting mixture was incubated under stirring at 37°C for 20 minutes. The resultant was subjected to ultrasonication for 15 seconds to completely
 5 disrupt the cells and the resultant was incubated at 37°C for 20 minutes, followed by centrifugation at 4°C at 12,000 rpm for 10 seconds. The supernatant was recovered and dialyzed against distilled water. The precipitated residue was subjected to the above-described operations several times to carry out extraction. The obtained extracted fluid was
 10 applied to a CM-Sepharose gel column equilibrated with 50 mM PKB (pH 6.0) and the column was washed with the same buffer. The adsorbed substances were then eluted with 1 M PKB (pH 6.0) and the eluted solution was dialyzed against distilled water overnight to remove salt, followed by concentrating the resultant with a collodion membrane,
 15 to obtain 1.5 mg of purified human medullasin.

Example 1

Preparation of Anti-human Medullasin Monoclonal Antibody

(1) Preparation of Hybridomas by Cell Fusion between Antibody-producing Cells and Myeloma Cells

20 The human medullasin extracted and purified from human granulocytes was emulsified with Freund's complete adjuvant and the resultant was subcutaneously administered to a BALB/c mice of 7 weeks old at a dose of 50 µg/mouse. Four weeks later, the mice were subjected to additional immunization by the same method as the first
 25 immunization. Seven days after the additional immunization, increase in the blood level of antibodies was confirmed. Another 7 days later, the antigen was intraperitoneally administered at a dose of 50 µg/mouse as

the final immunization. On the other hand, mouse myeloma P3-X63-Ag8-U1(P3U1) cells were passaged in DMEM supplemented with 20% fetus calf serum. Three days after the final immunization, spleen cells were collected from the mice and fused with P3U1 cells using polyethylene glycol 4000. After the cell fusion operation, the medium was changed to DMEM supplemented with 100 μ M hypoxanthine, 0.4 μ M aminopterin and 16 μ M thymidine (HAT medium), and the resultant was placed in wells of a 96-well microplate. By continuing the selective culture for 2 to 3 weeks, hybridomas between the spleen cells and the myeloma cells were obtained.

(2) Screening of Anti-human Medullasin Antibody-producing Hybridomas

The titers of the antibodies in the culture fluids of the hybridomas were determined by ELISA (enzyme-linked immunosorbent assay), thereby carrying out screening. That is, human medullasin was adsorbed on the wells of a microplate for ELISA, and the wells were blocked with 1% bovine serum albumin (BSA) solution in 10 mM phosphate-buffered saline (PBS) (pH 7.4). Fifty microliters of the hybridoma culture was added to each well and the resultant was left to stand for 1 hour. After removing the hybridoma culture and washing the wells, 100 μ l of 2 μ g/ml solution of peroxidase-labeled goat anti-mouse IgG Fc antibody in PBS was added to each well, and the resulting mixture was allowed to react at 37°C for 1 hour. After removing the enzyme-labeled antibody solution and washing the wells, 200 μ l of 0.1M phosphate citrate buffer (pH 4.6) containing 0.05% ABTS and 0.0034% hydrogen peroxide was added to each well to generate color, thereby selecting the anti-human medullasin antibody-producing hybridomas.

(3) Cloning of Antibody-Producing Cells and Preparation of Monoclonal Antibodies

Each of the cultures of the anti-human medullasin antibody-producing hybridomas was subjected to cloning by the limiting dilution method to finally obtain 4 kinds of monoclonal hybridoma. The hybridomas were separately administered to BALB/c mice intraperitoneally which mice preliminarily received pristane and the hybridomas were grown to obtain ascites each containing the monoclonal antibody. To the obtained ascites, 50% saturated ammonium sulfate was added to precipitate the antibody and the antibody was dissolved in PBS. The resulting solution was dialyzed against 50 mM Tris-HCl buffer (pH 7.8) containing 3 M NaCl and the resultant was applied to a Protein A-Sepharose CL4B Column (commercially available from Pharmacia). The adsorbed antibody was eluted with 0.1 M glycine-HCl buffer (pH 6.0) and the eluted solution was neutralized, followed by purification of antibody therefrom to obtain 4 kinds of monoclonal antibody, 3F03, 3G03, 2E04 and 1G12.

(4) Properties of Monoclonal Antibodies

Western Blotting

The antigen corresponding to the monoclonal antibodies was immobilized by Western blotting method.

First, medullasin from human granulocytes was subjected to SDS-polyacrylamide gel electrophoresis according to the method by Laemmli. The protein was transferred from the slab gel to a nitrocellulose sheet using an electrolyte buffer containing 25 mM Tris-(hydroxymethyl)aminomethane, 192 mM glycine and 20% methanol, at a voltage slope of 7 V/cm for 2 hours. Each lane on the nitrocellulose

sheet was cut and one of the sheets was subjected to protein staining with Amideblack and the other sheet was subjected to enzyme immunoassay as follows. That is, after blocking the sheet with 1% BSA/PBS, the mouse anti-human medullasin monoclonal antibody was added as a primary antibody, and then alkaline phosphatase-labeled goat anti-mouse IgG Fc antibody was added as a secondary antibody, and the resultant was allowed to react. After washing the sheet, a substrate solution containing 5-bromo-4-chloro-3-indolylphosphate ($0.7 \times 10^{-3}M$)/Nitroblue tetrazolium salt ($0.7 \times 10^{-3}M$) in 0.1M Tris-HCl buffer (pH 9.5) was added to generate color. By this, it was confirmed that all of the 4 mouse anti-human medullasin monoclonal antibodies recognize medullasin from human granulocytes.

Inhibition Assay

Human medullasin immobilized on the wells of a microplate for ELISA was reacted with a biotinylated first antibody in the presence of a non-labeled second antibody, and then avidin-conjugated peroxidase was reacted, followed by addition of a substrate solution to generate color, thereby carrying out inhibition assay. By this, with any combination of the monoclonal antibodies, the amount of the reacted biotinylated antibody was not changed. Therefore, it was confirmed that the 4 monoclonal antibodies recognize epitopes which are different each other.

Example 2

Immunoassay I of Human Medullasin

(1) Preparation of Beads on Which Monoclonal Antibody is Immobilized

After well washing polystyrene beads (6 mm diameter), the beads were immersed in 10 μ g/ml solution of mouse anti-human medullasin

monoclonal antibody (2E04) in PBS (pH 7.4) at 4°C for one day and the beads were washed with PBS, followed by blocking the beads with aqueous 1% BSA solution at 4°C for one day to obtain beads on which the monoclonal antibody was immobilized.

5 (2) Preparation of Peroxidase-labeled Monoclonal Antibody

To 1.0 mg/ml solution of mouse anti-human medullasin monoclonal antibody (2E04) in PBS, 0.1 ml of 10 mg/ml solution of N-(m-maleimide benzoic acid)-N-succinimide ester (MBS) in dimethylformamide was added and the mixture was allowed to react at 25°C for 30 minutes. The reaction mixture was applied to a Sephadex G-25 column and gel permeation chromatography was carried out using 0.1 M phosphate buffer (pH 6.0) to separate maleimide-bound monoclonal antibody and the non-reacted MBS.

On the other hand, to 1.0 mg/ml horse radish peroxidase (HRP) solution as an enzyme in PBS, 10 mg/ml solution of N-succinimidyl-3-(2-pyridylthio)propionate (SPDP) in ethanol was added and the mixture was allowed to react at 25°C for 30 minutes. The reaction mixture was applied to a Sephadex G-25 column and subjected to gel permeation with 10 mM acetate buffer (pH 4.5). The fractions containing pyridyl disulfide-bound HRP were collected and were about 10-fold concentrated in a collodion bag in ice. To the resultant, 1 ml of 0.1 M acetate-buffered physiological saline (pH 4.5) containing 0.1 M dithiothreitol was added and the resulting mixture was stirred at 25°C for 30 minutes to reduce the pyridyl disulfide group introduced into the HRP molecule, followed by gel permeation chromatography of the resulting mixture using Sephadex G-25 to obtain a fraction containing thiol-bound HRP.

Then the maleimide-bound monoclonal antibody and the thiol-

bound HRP were mixed and the mixture was concentrated to a protein concentration of 4 mg/ml in a collodion bag in ice. After leaving the resultant to stand at 4°C for one day, the resultant was subjected to gel permeation chromatography using Ultrogel AcA44 (commercially available from SEPRACOR) to obtain a peroxidase-labeled monoclonal antibody.

(3) Sandwich Enzyme Immunoassay of Human Medullasin

In a test tube, one bead on which the mouse anti-human medullasin monoclonal antibody (2E04) was immobilized, 50 µl of 2% BSA-containing PBS solution containing purified human medullasin (standard substance) in a concentration of 0, 1, 10, 100 or 200 ng/ml, and 350 µl of 2% BSA-containing PBS solution containing 0.2 µg/ml HRP-labeled mouse anti-human medullasin monoclonal antibody (2E04) were placed and the mixture was incubated at 37°C for 30 minutes. After removing the contents of the test tube by aspiration, the test tube was washed with physiological saline and 400 µl of 0.1 M phosphate citrate buffer (pH 4.6) containing 0.05% ABTS and 0.0034% hydrogen peroxide was added to each test tube. After incubating the mixture at 37°C for 30 minutes, 1 ml of 0.1N aqueous oxalic acid solution was added to each test tube to stop the enzyme reaction and the absorbance at 420 nm of the resulting solution was measured with a spectrophotometer. By plotting the measured absorbance with respect to the concentration of the standard substance, a calibration curve with good concentration dependence as shown in Fig. 1 was obtained using the same monoclonal antibody as the immobilized antibody and as the enzyme-labeled antibody.

Example 3

Immunoassay II of Human Medullasin

(1) Preparation of Beads on Which Monoclonal Antibody is Immobilized

After well washing polystyrene beads (6 mm diameter), the beads were immersed in 10 µg/ml solution of mouse anti-human medullasin monoclonal antibody (3F03) in PBS (pH 7.4) at 4°C for one day and the beads were washed with PBS, followed by blocking the beads with aqueous 1% BSA solution at 4°C for one day to obtain beads on which the monoclonal antibody was immobilized.

(2) Sandwich Enzyme Immunoassay of Human Medullasin

In a test tube, one bead on which the mouse anti-human medullasin monoclonal antibody (3F03) was immobilized, 50 µl of 2% BSA-containing PBS solution containing purified human medullasin (standard substance) in a concentration of 0, 1, 10, 100 or 200 ng/ml, and 350 µl of 2% BSA-containing PBS solution containing 0.2 µg/ml HRP-labeled mouse anti-human medullasin monoclonal antibody (2E04) were placed and the mixture was incubated at 37°C for 30 minutes. After removing the contents of the test tube by aspiration, the test tube was washed with physiological saline and 400 µl of 0.1 M phosphate citrate buffer (pH 4.6) containing 0.05% ABTS and 0.0034% hydrogen peroxide was added to each test tube. After incubating the mixture at 37°C for 30 minutes, 1 ml of 0.1N aqueous oxalic acid solution was added to each test tube to stop the enzyme reaction and the absorbance at 420 nm of the resulting solution was measured. By plotting the measured absorbance with respect to the concentration of the standard substance, a calibration curve with good concentration dependence as shown in Fig. 2 was obtained.

Example 4

Measurement of Medullasin in Clinical Samples by Enzyme Immunoassay

Samples of frozen blood collected from a normal individual and from a patient suffering from multiple sclerosis were thawed at room temperature and 10 μ l of each sample was added to 2 ml of PBS (pH 7.4), followed by uniformly mixing the mixture to prepare sample solutions.

5 To a test tube, 10 μ l of each sample solution was added and 40 μ l of 2.5% BSA-containing PBS (pH 7.4) was added to dilute the sample solution. To the test tube, one bead on which the mouse anti-human medullasin monoclonal antibody 2E04 or 3F03 was immobilized and 350 μ l of 2% BSA-containing PBS solution containing 0.2 μ g/ml HRP-labeled

10 mouse anti-human medullasin monoclonal antibody (2E04) were added and the resulting mixture was incubated at 37°C for 30 minutes. In exactly the same manner as in the above-described preparation of the calibration curves, washing, enzyme reaction, stopping of the reaction and measurement of absorbance at 420 nm with a spectrophotometer were

15 carried out, and the concentration of human medullasin was determined based on the calibration curve. As a result, the concentrations of human medullasin in the blood sample from the normal individual determined by Immunoassay I and Immunoassay II were 10.0 μ g/ml and 9.6 μ g/ml, respectively, and those in the blood sample from the patient suffering

20 from multiple sclerosis determined by Immunoassay I and Immunoassay II were 37.3 μ g/ml and 35.8 μ g/ml, respectively.

We claim:

1. An anti-human medullasin monoclonal antibody which specifically recognizes human medullasin.
2. The anti-human medullasin monoclonal antibody according to claim 1, which belongs to immunoglobulin class IgG.
3. The anti-human medullasin monoclonal antibody according to claim 1, which is originated from mouse.
4. A process for producing an anti-human medullasin monoclonal antibody comprising culturing hybridomas prepared by cell fusion between antibody-producing cells recovered from an animal immunized with human medullasin and myeloma cells, and recovering anti-human medullasin monoclonal antibody which specifically recognizes human medullasin from the culture in which said hybridomas are cultured.
5. An immunoassay comprising immobilizing human medullasin in a test sample by sandwiching said human medullasin between an anti-human medullasin monoclonal antibody immobilized on an insoluble carrier and a labelled anti-human medullasin monoclonal antibody by antigen-antibody reactions to form a complex, and quantifying said label in said complex.

A monoclonal antibody by which medullasin that is a kind of serine proteases existing in granulocytes, production process thereof and an immunoassay of human medullasin using the antibody are disclosed.

The monoclonal antibody specifically recognizes human medullasin. The process for producing the anti-human medullasin antibody comprises culturing hybridomas prepared by cell fusion between antibody-producing cells recovered from an animal immunized with human medullasin and myeloma cells, and recovering anti-human medullasin monoclonal antibody which specifically recognizes human medullasin from the culture in which the hybridomas are cultured. The immunoassay utilizes the anti-human medullasin antibody.

Figure 1

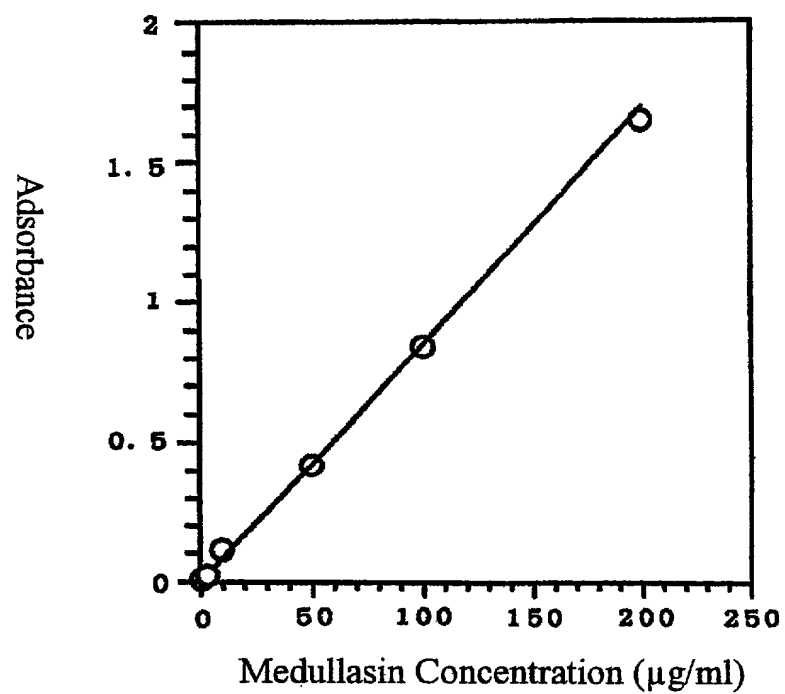
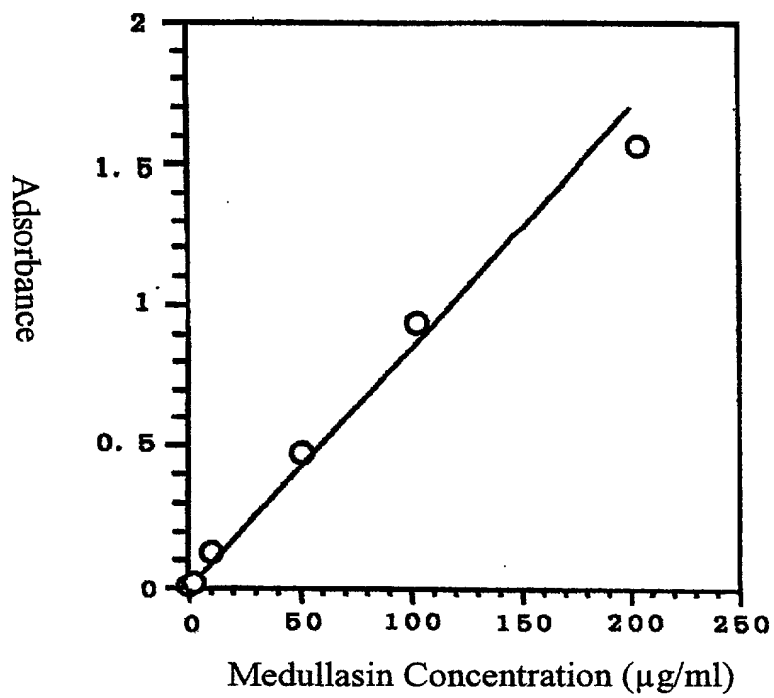


Figure 2



PLEASE NOTE:
YOU MUST
COMPLETE THE
FOLLOWING:



COMBINED DECLARATION AND POWER OF ATTORNEY FOR PATENT AND DESIGN APPLICATIONS

As a below named inventor, I hereby declare that: my residence, post office address and citizenship are as stated next to my name; that I verily believe that I am the original, first and sole inventor (if only one inventor is named below) or an original, first and joint inventor (if plural inventors are named below) of the subject matter which is claimed and for which a patent is sought on the invention entitled:

Insert Title: →

ANTI-HUMAN MEDULLASIN MONOCLONAL ANTIBODY, PROCESS FOR
PRODUCING THE SAME AND IMMUNOASSAY USING THE SAME

Fill in Appropriate
Information —
For Use →
Without
Specification
Attached:

the specification of which is attached hereto. If not attached hereto,

the specification was filed on _____ as
United States Application Number _____;
and amended on _____ (if applicable); and/or
the specification was filed on _____ as PCT
International Application Number _____; and was
amended under PCT Article 19 on _____ (if applicable)

I hereby state that I have reviewed and understand the contents of the above identified specification, including the claims, as amended by any amendment referred to above.

I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations, §1.56.

I do not know and do not believe the same was ever known or used in the United States of America before my or our invention thereof, or patented or described in any printed publication in any country before my or our invention thereof or more than one year prior to this application, that the same was not in public use or on sale in the United States of America more than one year prior to this application, that the invention has not been patented or made the subject of an inventor's certificate issued before the date of this application in any country foreign to the United States of America on an application filed by me or my legal representatives or assigns more than twelve months (six months for designs) prior to this application, and that no application for patent or inventor's certificate on this invention has been filed in any country foreign to the United States of America prior to this application by me or my legal representatives or assigns, except as follows.

I hereby claim foreign priority benefits under Title 35, United States Code, §119 (a)-(d) of any foreign application(s) for patent or inventor's certificate listed below and have also identified below any foreign application for patent or inventor's certificate having a filing date before that of the application on which priority is claimed:

Prior Foreign Application(s)

Priority Claimed

Insert Priority
Information: →
(if appropriate)

(Number)	(Country)	(Month / Day / Year Filed)	<input type="checkbox"/> Yes	<input type="checkbox"/> No
_____	_____	_____	<input type="checkbox"/>	<input type="checkbox"/>
_____	_____	_____	<input type="checkbox"/>	<input type="checkbox"/>
_____	_____	_____	<input type="checkbox"/>	<input type="checkbox"/>
_____	_____	_____	<input type="checkbox"/>	<input type="checkbox"/>

Insert Provisional
Application(s): →
(if any)

I hereby claim the benefit under Title 35, United States Code, §119(e) of any United States provisional application(s) listed below.

(Application Number)	(Filing Date)
_____	_____
_____	_____

All Foreign Applications, if any, for any Patent or Inventor's Certificate Filed More than 12 Months (6 Months for Designs) Prior to the Filing Date of This Application:

Insert Requested
Information: →
(if appropriate)

Country	Application Number	Date of Filing (Month / Day / Year)
_____	_____	_____
_____	_____	_____

I hereby claim the benefit under Title 35, United States Code, §120 of any United States and/or PCT application(s) listed below and, insofar as the subject matter of each of the claims of this application is not disclosed in the prior United States and/or PCT application in the manner provided by the first paragraph of Title 35, United States Code, §112, I acknowledge the duty to disclose information which is material to patentability as defined in Title 37, Code of Federal Regulations, §1.56 which became available between the filing date of the prior application and the national or PCT international filing date of this application:

Insert Prior U.S.
Application(s): →
(if any)

(Application Number)	(Filing Date)	(Status — patented, pending, abandoned)
_____	_____	_____
_____	_____	_____

I hereby appoint the following attorneys to prosecute this application and/or an international application based on this application and to transact all business in the Patent and Trademark Office connected therewith and in connection with the resulting patent based on instructions received from the entity who first sent the application papers to the attorneys identified below, unless the inventor(s) or assignee provides said attorneys with a written notice to the contrary:

Raymond C. Stewart (Reg. No. 21,066)
Joseph A. Kolasch (Reg. No. 22,463)
Bernard L. Sweeney (Reg. No. 24,448)
Charles Gorenstein (Reg. No. 29,271)
Leonard R. Svensson (Reg. No. 30,330)
Andrew D. Meikle (Reg. No. 32,868)
Joe McKinney Muncy (Reg. No. 32,334)
Donald J. Daley (Reg. No. 34,313)
John A. Castellano (Reg. No. 35,094)

Terrell C. Birch (Reg. No. 19,382)
James M. Slattery (Reg. No. 28,380)
Michael K. Mutter (Reg. No. 29,680)
Gerald M. Murphy, Jr. (Reg. No. 28,977)
Terry L. Clark (Reg. No. 32,644)
Marc S. Weiner (Reg. No. 32,181)
C. Joseph Faraci (Reg. No. 32,350)
John W. Bailey (Reg. No. 32,881)

Send Correspondence to: **BIRCH, STEWART, KOLASCH & BIRCH, LLP**
P.O. Box 747 • Falls Church, Virginia 22040-0747
Telephone: (703) 205-8000 • Facsimile: (703) 205-8050

PLEASE NOTE:
YOU MUST
COMPLETE
THE
FOLLOWING:
↓

I hereby declare that all statements made herein of my own knowledge are true and that all statements made on information and belief are believed to be true; and further that these statements were made with the knowledge that willful false statements and the like so made are punishable by fine or imprisonment, or both, under Section 1001 of Title 18 of the United States Code and that such willful false statements may jeopardize the validity of the application or any patent issued thereon.

Full Name of First or
Sole Inventor
Insert Name of
Inventor →
Insert Date This
Document is Signed

Insert Residence
Insert Citizenship →

Insert Post Office
Address →

Full Name of Second
Inventor, if any
see above

Full Name of Third
Inventor, if any
see above

Full Name of Fourth
Inventor, if any
see above

Full Name of Fifth
Inventor, if any
see above

GIVEN NAME	FAMILY NAME	INVENTOR'S SIGNATURE	DATE*
Yosuke	AOKI	<i>Yosuke Aoki</i>	Aug 24, 1999
Residence (City, State & Country)			CITIZENSHIP
Hino-shi, Tokyo JAPAN			JAPAN
POST OFFICE ADDRESS (Complete Street Address including City, State & Country)			
119 Tama-minamidaira Park Square, 19-2, Minamidaira 4-chome, Hino-shi, Tokyo 191-0041 JAPAN			
GIVEN NAME	FAMILY NAME	INVENTOR'S SIGNATURE	DATE*
Hideaki	SUZUKI	<i>Hideaki Suzuki</i>	Aug. 17, 1999
Residence (City, State & Country)			CITIZENSHIP
Adachi-ku, Tokyo JAPAN			JAPAN
POST OFFICE ADDRESS (Complete Street Address including City, State & Country)			
c/o DAINICHISEIKA COLOR & CHEMICALS MFG.CO., LTD. 9-4, Horinouchi 1-chome, Adachi-ku, Tokyo 123-8555 JAPAN			
GIVEN NAME	FAMILY NAME	INVENTOR'S SIGNATURE	DATE*
Kiyoshi	TAKAHASHI	<i>Kiyoshi Takahashi</i>	Aug. 17, 1999
Residence (City, State & Country)			CITIZENSHIP
Adachi-ku, Tokyo JAPAN			JAPAN
POST OFFICE ADDRESS (Complete Street Address including City, State & Country)			
c/o DAINICHISEIKA COLOR & CHEMICALS MFG.CO., LTD. 9-4, Horinouchi 1-chome, Adachi-ku, Tokyo 123-8555 JAPAN			
GIVEN NAME	FAMILY NAME	INVENTOR'S SIGNATURE	DATE*
Hisashi	KATSURAGI	<i>Hisashi Katsuragi</i>	Aug. 18, 1999
Residence (City, State & Country)			CITIZENSHIP
Adachi-ku, Tokyo JAPAN			JAPAN
POST OFFICE ADDRESS (Complete Street Address including City, State & Country)			
c/o DAINICHISEIKA COLOR & CHEMICALS MFG.CO., LTD. 9-4, Horinouchi 1-chome, Adachi-ku, Tokyo 123-8555 JAPAN			
GIVEN NAME	FAMILY NAME	INVENTOR'S SIGNATURE	DATE*
Residence (City, State & Country)			CITIZENSHIP
POST OFFICE ADDRESS (Complete Street Address including City, State & Country)			